Evaluation of Antimicrobial Activity of Different Aquatic Extracts Against Bacterial Isolates from UTI in Babylon Province, Iraq

Zainab Adil Ghani Chabuck¹, Baraa Hamid Hadi¹ and Nada Khazal Kadhim Hindi²

¹Department of Microbiology, College of Medicine, University of Babylon, Babylon Province, Iraq,
²PhD. Medical Microbiology/University of Babylon, Iraq, Head of basic and medical science\Nursing college/ Babylon University, Iraq.

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There are increments to the need of the usage of new non-chemical medicines, pharmaceuticals and health products. Plants are traditionally used as medicine over the world for their pharmacological values. They effectively used as anti-fungal, antiviral, anti-bacterial, anti-inflammatory, anti-diabetic, anti-oxidant, anti-ulcer, and antitussive. This study aims to investigate the effects of plant extracts (Glycyrrhiza glabra, Cuminum cyminum, Zingiber officinale, Origanum majorana and Petroselinum crispum) against different types of gram-positive and negative bacterial isolates. Antimicrobial activity of tested aqueous extracts by well-diffusion method against various bacterial isolates were done, to estimate their antibacterial activity. Results showed that the aquatic extracts of different plants produce a good antibacterial effect when compared with the synthetic antibiotic ciprofloxacin. As all bacterial isolates in this study where sensitive to these extracts with variable ranges of inhibition zones ranging from 18-32mm in diameter. Thus we can concluded that the aquatic extracts of Glycyrrhiza glabra, Cuminum cyminum, Zingiber officinale, Origanum majorana and Petroselinum crispum can be beneficial as treatment of UTI-causing bacteria.

Keywords: Glycyrrhiza glabra, Cuminum cyminum, Zingiber officinale, Origanum majorana, Petroselinum crispum.

Plants are rich sources of natural products that are extracted from them due to their content of active substances that used for centuries. These extracts can be used as antimicrobial activity to treat various diseases caused by pathogenic organisms in addition to their numerous pharamaceutical effects such as antibacterial, antifungal, antiparasitic and anticarcinogenic. The use of crude plant extracts in herbal medicine for treatment of diseases can be seen in different form like infusion or tincture. Bacterial resistance to different types of antibiotics is growing because of genetic variability and mutation therefore we must understand the genetic mechanisms of resistance to develop new drugs from plants that have natural products with safety and efficiency for human being.

Plant extracts widely used to reduce the bacterial growth of most infections. The antimicrobial activity of them dependent on their methods of extraction and chemical structures of plants. Glycyrrhiza glabra has important chemical components called licorice extracted from its roots that have medical benefits as antibacterial, antiviral and anti-ulcerative activity, it used for treatment of upper respiratory and liver diseases.

Cuminum cyminum has anticholesterol and antimicrobial activity and the active components
of it is cumin oil or cuminol, these extracts are important in stimulation of digestive system enzymes secretion to treat different types of diseases and used as bactericides against plant bacterial diseases 6,1.

Ginger (Zingiber officinale) has antimicrobial activity that can be used in treatment of bacterial infections through its aromatic and medicinal properties. Both ethanolic and aqueous extraction of Ginger have bactericidal effects on some types of pathogenic organisms 7,8. Petroselinum crispum extracts have antibacterial activity, damage bacterial cell wall and inhibit the bacterial growth in addition to their cytotoxic activity at higher concentration 9.

Many researches found the Origanum majorana has an essential oils that affect the bacterial growth through altering cell membrane permeability, and causing a distortion of the membrane structure, also the extracts of this plant has agricultural, pharmaceutical and interfere with cosmetic industries 10.

The antimicrobial activity of plant extracts have been compared with the effects of different types of antibiotics through measuring the inhibition zones of bacterial growth. Ciprofloxacin has a wide antibacterial spectrum against the pathogenic gram-positive and -negative bacteria that caused different types of urinary tract, respiratory tract, skin infections and other types of infections through inhibition the action of enzymes that interfere with bacterial replication 11.

**Aim**

This study aims to investigate the action of plant extracts (Glycyrrhiza glabra, Cuminum cyminum, Zingiber officinalis, and Petroselinum crispum) against a number of positively- and negatively-gram stained bacterial isolates and compare them with the effects of ciprofloxacin.

### MATERIAL AND METHODS

**Extraction Methods**

Plants were collected from the market. Aqueous extracts were prepared by soaking 30 grams of powder in 100 ml of distilled water, leave them to stand for 72 hours, and sterilized by using Millipore 0.45 filter paper. 50% concentration of the extract was obtained by this way 12.

**Bacterial Isolates**

An overall of eight negatively-gram stained and three positively-gram stained bacterial isolates were collected from urine clinical samples (UTI) and used in this study. These bacterial isolates were; S. saprophyticus, S. epidermidis, S. aureus, P. fluosescence, P. aeruginosa, E. aerugenese, E. coli, K. pneumoniae, P. vulgaris, Acinetobacter, Proteus mirabilis. All these bacterial isolates were cultivated for activation and re-cultured as three successive times on nutrient agar plates. Identification and diagnosis of these isolates were applied by conventional biochemical procedures 13.

**Agar Well Diffusion Assay for In vitro Testing the Antimicrobial Activity (14)**

Bacterial growths of Loopfull from each bacterial isolates were taken and applied into nutrient broth, incubation at 37°C for 18 hours. Normal saline was used to dilute the bacterial suspensions to adjust the turbidity and comparing with standard McFarland tube number 0.5. Cotton swab dipped into the tube of suspension and streaked Mueller-Hinton agar plates, left them to dry for 5-15 minutes at room temperature. Four wells of about 5 mm in diameter were made in the media by cork borer, 0.1 ml of each extracts were added. Then incubation overnight at 37°C. Zone of inhibition was measured, disks of ciprofloxacin were add in the center of agar plate to compare the results of bacterial inhibition 13.

### RESULTS AND DISCUSSIONS

The evaluation of bacterial inhibitory activity of aquatic extracts of Glycyrrhiza glabra was processed using the agar diffusion test, as in Figure (1). This figure shows that Glycyrrhiza glabra exhibited an inhibitory activity against all the bacterial isolates of the study with inhibition zones ranging from 18-25 mm, with the largest inhibition against E. coli (25 mm) followed by Pseudomonas aerogenosa and Staphylococcus saprophyticus (24 and 23 mm) respectively. Lowest inhibition against Klebsiella pneumonia and Enterobacteria aerogenes (18 mm) for each.

Figure (2) shows the inhibitory activity of aquatic extracts of Cuminum cyminum against different bacterial isolates in the study. Results demonstrated that inhibition zones were the
Fig. 1. Antibacterial activity of *Glycyrrhiza glabra* (ÜÑÞ ÖxeÖ) against bacterial isolates

Fig. 2. Antibacterial activity of *Cuminum cyminum* (błæä) against bacterial isolates

Fig. 3. Antibacterial activity of ginger(ØülÉüá) against bacterial isolates
Chabuck et al.: Antimicrobial Activity of Aquatic Extracts

**Fig. 4.** Antibacterial activity of *Petroselinum crispum* (ÉPÎæäÓ) against bacterial isolates

**Fig. 5.** Antibacterial activity of *Origanum majorana* (ãÑÏÞæÔ) against bacterial isolates

**Fig. 6.** Antibacterial activity of Ciprofloxacin against bacterial isolates

J PURE APPL MICROBIOL, 12(2), JUNE 2018.
same (20mm) against all the tested bacterial isolates. While the diameter of inhibition zones produced by Ginger were ranging from 23mm against Enterobacter aerogenes to 18mm with Klebsiella pneumonia; and 22mm against most of isolates, figure (3).

The antibacterial effect of Petroselinum crispum exhibited as a high inhibition level as compared with other extracts, as the inhibition zone diameters range from 32-28mm; higher effects against Klebsiella pneumonia, Enterobacter aerogenes and Acinetobacterspp (32mm), followed by 30mm as an inhibition against 6 bacterial isolates, and the lowest one (28mm) with Proteus mirabilis and Proteus vulgaris (4).

Results in table (5) revealed the antibacterial inhibitory effect of Origanum majorana Gram positive and negative bacteria with the highest inhibition 32 mm against E. coli, 30 mm against seven bacterial isolates, while the least inhibition against Pseudomonas aeruginosaand Pseudomonas fluorscence(28) and 27mm respectively. In contrast to all these aquatic extracts, Ciprofloxacine antibiotic is used as a synthetic antibacterial reagent; figure (6). This showed resistance of bacteria to this reagent; especially Pseudomonas aeruginosaand Pseudomonas fluorscence 0 mm, while the largest effect were only reach to about (17 mm) with staphylocci and E.coli, smallest zones were 12 mm with Proteus spp.

Extract of Licorice root has been used traditionally for a long period od history of liquorice root extract. It is considered to be the most commonly used of the folkmedicines in Asia. Its structure is a saponin compound16,17. Thus due of the existence of these different secondary metabolites such as flavonoids, alkaloids, and saponins in its aquatic-root extract, it exhibits potent antibacterial activity18,19, also it has a role as antiviral agent against many viruses20,21.

In vitro works had demonstrated that aquaticand ethanolic extracts of liquorice display an inhibitory action on the cultures of Strept. pyogenesand Staph. aureus(22), and many pathogenic organisms23.

In vitro testing of the extracts from liquorice root was carried out against3 bacterial isolates by the use of agar diffusion method, that showed various antibacterial activities. The inhibitory antibacterial properties of this extract to prevent the growth ofS. flexneri, Shigella sonnei, S. paratyphi BSalmonella typhi and ETEC E. coli was revealed24.

Glycyrrhizin had been studied by Alonso and Tratado25, as it is regularly used amongthe orally administered foods, and it obstructs the growth of some bacterial sorts, also can prevent dental caries formation, it demonstrated antimicrobial effects against both Gram-positive and -negative bacteria by Gupta et al26.

Cumin spice is used popularly as a traditional therapy, for the reason that aromatic substances are present in this herb, it is prescribed for the mild digestive sicknesses, and used in the treatment of diseases caused by K. pneumoniae; especially the essential oil of its seeds that had displayed to have a significant in vitro antibacterial activity26,27. Cuminaldehyde and para-cymene are the main two active components of cumin; Cuminaldehyde has made known divers activities like antifungal, antibacterial, anti-diabetic and anti-platelet28. Antimicrobial effect of both aqueous and oil extracts of it, had judged against a wide collection of valuable, pathogenic microbial strains of both gram-positive and -negative, as it is reported to obstruct Salmonella spp, E.coli, Aspergillus niger and Bacillus cereus growth29,30.

Alcoholic extract and oil of Cumin seedshave ability to inhibit growth of Klebsiella pneumoniae, especially its clinical isolates, with improvement in the morphology of cells, decrement of urease activity and impeding capsule expression. Additionally, have capability to prevent properties of Streptococcus pyogenes and Streptococcus mutansin biofilm formation. Moreover the cumin extracts has anti-fungal properties against human, soil, food and animal pathogens, yeasts, mycotoxin and aflatoxins producers31,32,33.

Ginger exhibited a robust antibacterial and to a lessdegree antifungal properties. Main active ingredients of ginger have been studied in vitro to inhibit multiplication and growth of colon bacteria; as ginger can counteract bacterial
ability to produce flatulence due to fermentation of undigested carbohydrates. It constrains growth of *Staphylococci*, *Salmonella*, *Proteus sp.*, *E. coli* and *Streptococci* (34). Gingerol, Ingenol and Shogaol, isolated from ginger rhizome are the main active components that exhibited antiviral and antibacterial activity against many pathogens; as *M. avium* and *M. tuberculosis* in vitro, *H. pylori* and many periodontal bacteria (35,36,37).

Hindi *et al.* (12) showed that the aquatic extract of ginger give a significant influence to inhibit most of the studied sixteen bacterial isolates both gram positive and negative, also in a study of Suhad and her coworkers displayed that the ginger aquatic extract gave the peak effect against *Staphylococcus aureus*, *Streptococcus pyogenes*, *Proteus vulgaris*, and *Klebsiella pneumonia*. A number of antibiotics like chloromphenicol, erythromycin, gentamicin, nalidixic acid and trimethoprim were used as a comparison with this extract of ginger, using antibiogram test. Aa resultstheinstituted that the inhibitory action of ginger was superior than that to Gentamicin and Chloromphenicol against *Proteus vulgaris*, *Klebsiellapneumoniae, Staphylococcus pyogenes* and *Streptococcus pyogenes* (38).

Various compounds from different phytochemical categories have been identified in Parsley. Also, different pharmacological activities have been attributed to it. In Iran Parsleys seeds or leaves has many usages for renal system, as inflammation, kidney stone, renal and urinary tract disease, and urinary tract infections (39,40,41). Also, extracts from its leaves are widely used in Serbia as an antimicrobial agent (42).

As Parsley leaves and stems possess antibacterial activity on *B. subtilis* and *E. coli*. Both hot and cold water extract from them demonstrated antibacterial activity against *pseudomonas aeruginosa, S. aureus, S. epidermard S. pyogenes*, with higher inhibition zone in hot water extract (43). Leaf extract showed higher cell damage on both bacteria with higher activity with methanol extract, Coumarins are responsible components for this activity (44,45).

Microbial analysis carried out by Nessrien *et al.* (46) and Gutierrezet al. (47) showed that, essential oils of marjoram has antimicrobial properties, as it is rich in phenolic compound that being particularly active as antimicrobials against both bacteria and fungi.

The essential oils from its leaves showed antibacterial effects on various bacteria (*Bacillus cereus, E. coli, Staphylococcus aureus*, *Proteus spp.*, *Enterobacter spp.*, *Klebsiella* spp., *Acinetobacter* spp. and *Pseudomonas* spp.) in agar diffusion assay. Also the ethanol and water extracts of majorana have shown antimicrobial activity against both gram positive and negative bacteria and its possible food applications by minimum inhibitory concentration estimation (48).

Faroqui and Sreeramu (49) have reported the antimicrobial activity of majorana against *Bacillus anthracis, Proteus vulgaris, Salmonella spp.*, *Streptococcus agalactiae, Streptococcus spp.* and *Aspergillus fumigatus*. Shahidi (50) screened some used medical Iranian traditional plants for antibacterial properties against two *E. coli* strains and found that Majorana showed anti-*E. coli* activity. Leelavathi (51) has conducted a comparative study of the antibacterial activity of crude extract of *in vivo* and *in vitro* leaves of majorana against *Staphylococcus aureus* and reported that *in vitro* leaf extract showed better antibacterial activity.

At present, most pathogenic bacteria develop an antibiotic resistance. Thus to overwhelm this frightening problem, it is an urgency to discover a number of novel active compounds. Organic solvents and water are used to prepare extracts from these spices which are biologically active compounds, and can be applied for the synthesis of potent drugs. Thus spices, that considered as ausual components of our routine food preparations, which may give defense to a certain extent against bacterial pathogens the natural enemies.

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